



ORIGINAL ARTICLE

Serum and tissue angiogenin in patients with endometrial hyperplasia

Mohamed Abelsalam Mohamed ^{a,*}, Naglaa F. Abraheem ^b

^a *Obstetrics and Gynecology Department, Benha Faculty of Medicine, Benha, Egypt*

^b *Biochemistry and Molecular Biology Department, Benha Faculty of Medicine, Benha, Egypt*

Received 11 May 2010; accepted 19 July 2010

Available online 11 November 2010

KEYWORDS

Endometrial hyperplasia;
Atypia;
Angiogenin

Abstract Objective: To evaluate angiogenin levels in both tissue and serum of patients with endometrial hyperplasia with and without atypia.

Methods: Sixty women were classified according to the histopathological findings of endometrium into three groups. The control group consisted of 20 women with normal non-hyperplastic endometrium. The second group included 20 women diagnosed as complex endometrial hyperplasia without atypia. The third group included 20 women diagnosed as complex endometrial hyperplasia with atypia. Serum and tissue angiogenin were measured by enzyme immunoassay (EIA) technique and confirmed in tissues with Western Blotting (WB) technique.

Results: There was a statistically significant increase in serum and tissue angiogenin levels of endometrial hyperplasia groups compared to those of control group ($P < 0.001$). Serum and tissue angiogenin levels were with a statistically significant higher ($P < 0.001$) in group III compared to group II. The sensitivity of serum angiogenin to detect the potential possibility of endometrial hyperplasia with atypia in endometrial hyperplasia patients was 100%, specificity 85%, positive predictive value 86.9%, negative predictive value 100%, positive likelihood ratio 6.6%, negative likelihood ratio 0% and accuracy 91.7%.

Conclusion: Elevated levels of serum angiogenin in endometrial hyperplasia could assist in determining which patients are at high risk for atypical change requiring aggressive treatment.

© 2010 Middle East Fertility Society. Production and hosting by Elsevier B.V. All rights reserved.

* Corresponding author. Tel.: +20 0123723084.

E-mail address: mahamadsalam@hotmail.com (M.A. Mohamed).



1. Introduction

Endometrial hyperplasia (EH) is a non-invasive proliferation of the lining of the uterus that results in a spectrum of glandular alteration (1). EH is classified primarily as either simple or complex, based on the degree of architectural complexity as seen by glandular crowding (with back-to-back crowding in the case of complex hyperplasia), and with or without cytological atypia; that is, nuclear irregularity, such as the loss of axial

polarity; rounded, stratified nuclei and prominent nucleoli (2). The wide range of histomorphological presentation of endometrial hyperplasia is accompanied by high intra and inter observer variability in diagnostic classification (3). Previous studies have shown that only 10–20% of EH progress to carcinoma when left untreated (4).

The lack of criteria that could accurately predict the disease outcome may have been an important cause of over and under treatment and hence the need of establishment of a new classification composed of three groups: endometrial hyperplasia (EH), endometrial intraepithelial neoplasm (EIN) and endometrial carcinoma (3). EIN is defined as neoplastic focal lesion with cytological features of crowded gland architecture and a volume percentage less than 55%, with a minimum size of 1 mm and careful exclusion of mimics (5,6). It's important to characterize high or low risk groups before initiation of therapy, because about 1–28% of hyperplasia progress to carcinoma, depending on the degree of severity (7).

Angiogenesis, the process of new blood vessel growth, plays an essential role in normal physiological processes, such as development and reproduction. However, pathological angiogenesis occurs in many angiogenesis dependent diseases such as tumours and other non-neoplastic diseases (8). Angiogenin (ANG) is the first human tumour cell-derived protein with *in vivo* angiogenic activity. ANG, a heparin binding 14.1 kDa single chain polypeptide, was initially isolated from supernatants of colon cancer cells and was found to be a member of the pancreatic ribonuclease superfamily (9).

Endometrial adrenomedullin, microvessel density and area of venules, which are tissue markers of angiogenesis, increase in a stepwise manner from normal, simple or complex hyperplasia with or without atypia to grade I adenocarcinoma (10).

Treatment of EH depends on the patient's age, fertility desire and the type of hyperplasia. Progestagens are still the most commonly used medical treatment modality in these patients. Response rates are higher for cases without atypia. In selected cases, hysterectomy may be performed as a definitive treatment modality (11).

The aim of the present study was to measure ANG levels in both tissue and serum of patients with endometrial hyperplasia with or without atypia.

2. Materials and methods

2.1. Patients

This case control study was performed on sera and endometrial biopsies from 60 women attending the University Hospital in Benha between January 2006 and January 2009. The study was approved by the local institutional review board and all women gave informed consent before enrollment in this study. They were classified according to the histopathological findings of the endometrium into three groups. The control group consisted of 20 women with normal non-hyperplastic endometrium. They were selected from perimenopausal women attending the out patient clinic for gynecological consultation. The second group included 20 women who complained of perimenopausal bleeding and diagnosed as complex endometrial hyperplasia without atypia. The third group included 20 women who complained of perimenopausal bleeding and diagnosed as complex endometrial hyperplasia with atypia. None

of the women had received preoperative hormonal therapy. Patients with EH underwent total abdominal hysterectomy.

Five milliliters of venous blood samples were collected before any treatment and preoperatively for all women and one week postoperatively in hyperplasia groups. The endometrial specimens had been obtained by dilatation and curettage. Blood clots, debris and muscle tissue were rapidly dissected from the endometrial tissue. One part of the endometrial specimen was sent for pathologic examination and the other part was washed with ice-cold saline, immediately frozen and stored at -70°C until analysis. After stripping away blood clots, debris and muscle tissue, the samples were thawed on ice, placed in 10 volume of ice-cold cell lysis buffer (pH 7.8, containing 100 mmol/l $\text{K}_2\text{HPO}_4\text{-KH}_2\text{PO}_4$, 1 mmol/l DTT, 2 mmol/l EDTA, 1% Triton X100, and 0.75 $\mu\text{g/ml}$ leupeptin) and homogenized. The cell lysate was centrifuged and the supernatant recovered and stored at -70° until analysis. The total protein in the prepared supernatant was measured by Bradford method using bovine serum albumin as a calibrator.

2.2. Determination of angiogenin by EIA

Quantitative determination of human angiogenin concentration in serum and tissues were done by a solid phase EIA (Quantikine, R&D System, USA). The assay employs the quantitative sandwich enzyme immunoassay technique (12).

2.3. Determination of ANG by Western Blotting (WB) technique

Twelve percentage of sodium dodecyl sulfate (SDS) polyacrylamide gel electrophoresis was used to separate 20 μg sample protein of tissue supernatant. The gels were transferred to nitrocellulose (NC) filters in Tris-glycine buffer (25 mM Tris, 192 mM glycine, 20% methanol, pH 7.4) for 1 h at 60 V. NC sheets were washed and the unoccupied binding sites were saturated with blocking solution (Chromogenic Western Blotting kit, Biorad-Roche Diagnostics, GmbH, Germany) for 1 h at 37°C . The sheets were then incubated with 0.1 mg/ml of either antihuman angiogenin monoclonal antibody (MOAB) (R&D Systems, USA) or normal mouse IgG serum (negative control) overnight at 4°C . The membranes were washed with Tris buffer saline (TBS) (50 mM Tris, 150 mM NaCl, pH 7.5). The antibodies that bound to the NC membrane were visualized by incubation with anti-mouse IgG-alkaline phosphatase conjugate for 90 min at room temperature (RT). Finally, the filters were incubated with alkaline phosphatase substrate solution (5-bromo-4-chloro-3-indolyl phosphate/nitroblue tetrazolium in 0.1 M Tris buffer) at RT until the developed bands were of desired intensity. By comparing the resulting developed NC with others in which normal mouse IgG serum was substituted for ANG MOAB, the ANG band was identified.

2.4. Statistical analysis

Anova test was used for statistical comparison of age and parity among the study groups. The non-parametric Kruskal-Wallis, and Mann-Whitney U rank sum tests were used for the statistical comparison of the ANG median value in the study groups. Spearman's rank correlations test was used to correlate serum and tissue ANG. Paired *t* test was used for the statistical comparison of the serum ANG values before

Table 1 Mean values + SD of age (years) and parity in the study groups.

Groups	Age (years)	Parity
– Control (<i>n</i> = 20)	45.8 ± 3.9	2.6 ± 1.2
– Endometrial hyperplasia without atypia (<i>n</i> = 20)	45.3 ± 2.1	2.7 ± 1.4
– Endometrial hyperplasia with atypia (<i>n</i> = 20)	46.1 ± 2.2	2.9 ± 1.2

P-value for age = 0.672.

P-value for parity = 0.750.

and after surgery. Significance was accepted for $P < 0.05$. All analyses were performed using the Statistical Package for the Social Sciences (SPSS, Chicago, Illinois) on an IBM personal computer.

3. Results

There was no statistically significant difference among the three study groups regarding age and parity (Table 1).

There was a statistically significant difference in serum and tissue ANG levels between EH groups and control groups ($P < 0.001$). Serum and tissue ANG levels were with a statistically significant higher ($P < 0.001$) in group III compared to group II (Table 2). There was a statistically significant reduction ($P < 0.001$) in serum ANG after operation in endometrial hyperplasia groups (Groups II and III) (Table 3). Using serum ANG 160 pg/ml as a cutoff, all the control women have serum ANG less than the cutoff value, 15% of the EH without atypia group and 100% of the EH with atypia group had serum ANG above the cutoff value which is a statistically significant difference ($P = 0.001$) (Table 4).

The sensitivity of serum ANG to detect the potential possibility of EH with atypia in EH patients was 100%, specificity 85%, positive predictive value 86.9%, negative predictive value 100%, positive likelihood ratio 6.6%, negative likelihood ratio 0% and accuracy 91.7%.

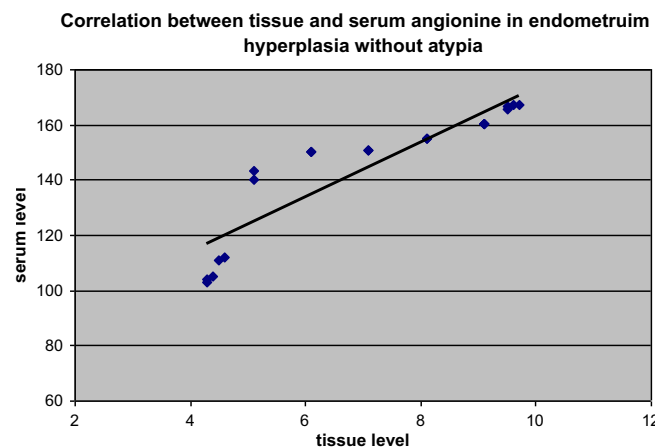
There was a significant positive correlation between serum ANG and tissue ANG in group II ($r = 0.9966$, $P < 0.0001$) (Fig. 1) and group III ($r = 0.9955$, $P < 0.0001$) (Fig. 2).

Table 3 Serum ANG concentrations (pg/ml) before and after the surgery in patients groups.

	Before surgery	After surgery	<i>P</i> -value
Group II	133.2 ± 9.1	79.8 ± 3.2	< 0.001
Group III	199.8 ± 8.1	89.1 ± 3.1	< 0.001

Table 4 Positivity rate of serum ANG concentrations in the study groups.

	Group I	Group II	Group III	<i>P</i> -value
Serum ANG				
> 160 pg/ml	0 (0%)	3 (15%)	20 (100%)	$P = 0.001$
< 160 pg/ml	20 (100%)	17 (85%)	0 (0%)	

**Figure 1** Correlation between tissue and serum angiotensin in endometrial hyperplasia without atypia.

4. Discussion

It was reported that the expression of ANG was upregulated in various types of human cancers, including breast, cervical, endometrial, ovarian and colon cancer (13). This indicates a close relationship between ANG and tumour development by

Table 2 Serum and tissue angiotensin concentrations with study groups.

	Group I (<i>n</i> = 20)	Group II (<i>n</i> = 20)	Group III (<i>n</i> = 20)
<i>Serum ANG (pg/ml)</i>			
Mean + SEM	65.26 ± 5.36	143.95 ± 5.16	201.42 ± 5.65
Range	41.1–101.3	103.2–167.5	163.7–237.1
Mean rank (median)	10.5 (55.65)	30.9 (150.9) ^a	50.1 (199.85) ^{a,b}
Kruskal–Wallis test	51.43, $P < 0.0001$		
<i>Tissue ANG (pg/mg protein)</i>			
Mean + SEM	1.93 ± 0.24	7.025 ± 0.47	12.535 ± 0.53
Range	0.95–4.8	4.3–9.7	9.9–16.1
Mean rank (median)	10.8 (1.45)	30.3 (7.1) ^a	50.5 (12.25) ^{a,b}
Kruskal–Wallis test	51.81, $P < 0.0001$		

Mann–Whitney U test:

^a Significant difference compared to the control group, $P < 0.001$.

^b Significant difference compared to endometrial hyperplasia without atypia group, $P < 0.001$.

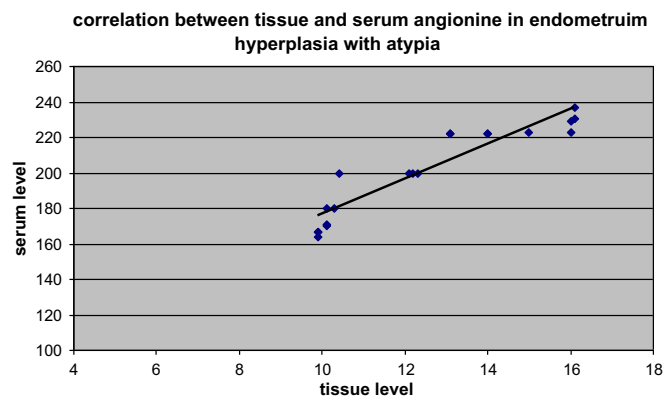


Figure 2 Correlation between tissue and serum angiogenin in endometrial hyperplasia with atypia.

stimulating both angiogenesis and cancer cell proliferation (14). Angiogenesis in EH was studied several years ago, Erdem et al. (15) demonstrated higher expression of vascular endothelial growth factor (VEGF) in EH than proliferative endometrium. Nunobiki et al. (10) demonstrated the role of adrenomedullin and microvessel density in hyperplastic endometrium. Consistent with these previous reports, the present results demonstrated that serum and tissue ANG concentrations measured by EIA and confirmed by WB technique, were significantly elevated in EH with atypia patients as compared with EH without atypia and female controls.

In the present study, the source of elevated serum ANG in the sample from the patient groups was not definitively elucidated. However, the obtained positive correlation between serum and tissue ANG could suggest that the source of serum ANG was the hyperplastic endometrial tissue. Because many types of other cells such as peripheral blood cells, vascular endothelial cells, fibroblasts and normal endometrial epithelial cells express ANG mRNA (16), these cells could be the source of serum ANG in EH group as well as in controls. However, it is probable that EH can contribute to the increased serum ANG concentration. This hypothesis is supported both by the previous findings that the endometrial hyperplasia tissue is angiogenic (17) and by the present findings of a reduction in serum ANG concentration after operative resection to approach the level as the in control group. The present result revealed that the median serum ANG in EH with atypia patients was 1.3 times that of EH without atypia patients. More over the median tissue ANG in EH with atypia was 1.7 times that for EH without atypia. This means that ANG is an important landmark for atypical change in EH. This result was previously documented in ovarian neoplasm (18), cervical neoplasm (19) and gestational trophoblastic tumour (20). In addition, the obtained data showed that in the EH with atypia group, the rate of patients with elevated serum ANG concentration more than the cutoff (160 pg/ml) was 100% while none of the control and only 15% of EH without atypia patients were positive for serum ANG ($P = 0.001$). This could mean that elevated levels of serum ANG in EH could assist in determining which patients are at high risk for atypical change and require aggressive treatment.

5. Conclusion

The present study demonstrates that serum ANG could be useful as a diagnostic marker for atypical EH and could predict

the atypical changes in cases of EH. From the clinical point of view, serum ANG level above 160 pg/ml can be used as a cutoff value for determining the patients who may need surgical treatment.

6. Conflict of interest statement

None declared.

References

- (1) Mazur MT, Kurman RJ. Diagnosis of endometrial biopsies and curettings: a practical approach. New York, NY: Springer Publishing Company; 2005.
- (2) Silverberg SG, Kuman RJ, Nogales F, et al. Tumours of the uterine corpus: epithelial tumours and related lesions. In: Tavassoli FA, Devilee P, editors. Pathology and genetics: tumours of the breast and female genital organs. Lyon, France: IARC Press; 2003. p. 221–32.
- (3) Baak JPA, Mutter GL. EIN and WHO 94. J Clin Pathol 2005;58: 1–6.
- (4) Orbo A, Baak JPA, Kleivan I, Lysne S, Prytz PS, Broeckerart MAM, et al. Computerized morphometrical analysis in endometrial hyperplasia for the prediction of cancer development a long term retrospective study from northern Norway. J Clin Pathol 2000;3:697–703.
- (5) Baak JP, Van Diermen B, Steinbakk A, Janssen E, Skaland I, Mutter GL, et al. Lack of PTEN expression in endometrial intraepithelial neoplasia is correlated with cancer progression. Hum Pathol 2005;36:555–61.
- (6) Mutter GL. The endometrial collaborative group. Endometrial intraepithelial neoplasia (EIN): will it bring order to chaos? Gynecol Oncol 2000;76:287–90.
- (7) Baak JP, Mutter GL, Robboy S, Van Diest PJ, Uytterlinde AM, Orbo A, et al. The molecular genetics and morphometry-based endometrial intraepithelial neoplasia classification system predicts disease progression in endometrial hyperplasia more accurately than the 1994 World Health Organization classification system. Cancer 2005;103:2304–12.
- (8) Folkman J. Angiogenesis: an organized principle for drug discovery? Nat Rev Drug Discov 2007;6:273–86.
- (9) Shimoyama S, Yamasaki K, Kawahara M, Kaminishi M. Increased serum angiogenin concentration in colorectal cancer is correlated with cancer progression. Clin Cancer Res 1999;5:1125.
- (10) Nunobiki O, Nakamura M, Taniguchi E, Utsunomiya H, Mori I, Tsubota Y, et al. Adrenomedullin, BCL-2 and microvessel density in normal, hyperplastic and neoplastic endometrium. Pathol Int 2009;59(8):530–6.

- (11) Galtekin M, Diriball K, Dursum P, Ayhan A. Current management of endometrial hyperplasia and endometrial intraepithelial neoplasia (EIN). *Eur J Gynecol Oncol* 2009;30(4):396–401.
- (12) Miyake H, Hara I, Yamanaka K, Gohij K, Arakawa S, Kamidono S. Increased angiogenin expression in the tumour tissue and serum of urothelial carcinoma patients is related to disease progression and recurrence. *Cancer* 1999;86(2):316–24.
- (13) Yoshioka N, Wang L, Kishimoto K, Tsuji T, Hu GF. Therapeutic target for prostate cancer based on angiogenin-stimulated angiogenesis and cancer cell proliferation. *Proc Natl Acad Sci USA* 2006;103:14519–24.
- (14) Xiangwei G, Zhengping X. Mechanisms of action of angiogenin. *Acta Biochim Biophys Sin* 2008;40:619–24.
- (15) Erdem O, Erdem M, Erdem A, Memis L, Akyol G. Expression of vascular endothelial growth factor and assessment of microvascular density with CD 34 and endoglin in proliferative endometrium, endometrial hyperplasia and endometrial carcinoma. *Int J Gynecol Cancer* 2007;17(6):1327–32.
- (16) Shimoyama S, Gansauge F, Gansauge S, Negri G, Oohara T, Beger HG. Increased angiogenin expression in pancreatic cancer is related to cancer aggressiveness. *Cancer Res* 1996;56:2703.
- (17) Abulafia O, Triest WE, Shere DM, Hansen CC, Ghezzi F. Angiogenesis in endometrial hyperplasia and stage I endometrial carcinoma. *Obstet Gynecol* 1995;86(4 PT1):479–85.
- (18) Brustmann H, Riss P, Naude S. The relevance of angiogenesis in benign and malignant epithelial tumours of the ovary: a quantitative histologic study. *Gynecol Oncol* 1997;67:20–6.
- (19) Bodner-Adler B, Hefler L, Bodner K, Leodolter S, Frishchmuth K, Kainz C, et al. Serum levels of angiogenin (ANG) in invasive cervical cancer and in cervical intraepithelial neoplasia (CIN). *Anticancer Res* 2001;21(1B):809–12.
- (20) Shaarawy M, El-Mallah SY, Sheiba M. Angiogenin and gestational trophoblastic tumours, a promising prognostic marker. *Clin Chem Lab Med* 2003;41(3):306–10.